

18-MALONYLOXY-9-EPI-ENT-ISOPIMAROL, A DITERPENE FROM *CALCEOLARIA GLANDULOSA**

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Abstract—A new pimarane diterpene, 18-malonyloxy-9-*epi-ent*-isopimarol, was isolated from the aerial parts of *Calceolaria glandulosa*. The structure of the new compound was established by spectroscopic evidence, chemical transformations and X-ray analysis.

INTRODUCTION

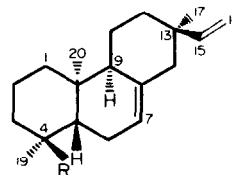
Calceolaria is a genus mostly of medium size herbs widespread in South America, 10% of which are endemic to Chile [2]. In continuation of our investigation of the genus *Calceolaria* [1, 3, 4] we have now examined *C. glandulosa*, an uncommon taxon distributed in the coastal hills of central Chile. This paper deals with the structural determination of the major compound, a new pimarane diterpene named 18-malonyloxy-9-*epi-ent*-isopimarol (1), isolated from the aerial parts of the plant.

RESULTS AND DISCUSSION

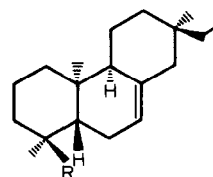
The chloroform extract of the fresh aerial parts of *C. glandulosa* was subjected to CC on silica gel using increasing proportions of ethyl acetate in petrol. Compound 1, obtained from the intermediate fractions as a colourless oil, showed a molecular formula $C_{23}H_{34}O_4$ by mass spectrometry ($[M]^+$ at m/z 374) and ^{13}C NMR, and its IR spectrum exhibited carboxyl, ester and olefinic group absorptions. The acidic nature of this substance was evident from its characteristic chromatographic behaviour and because, by reaction with ethereal diazomethane, the methyl ester 1a was easily obtained (see Experimental and Table 1).

The 1H NMR spectrum of 1 showed the presence of a vinyl group (δ 5.84, *dd*, H-15, 4.96, *dd*, H-16c; 4.94, *dd*, H-16t), a primary ester group (δ 3.95, *d*, H-18, 3.78, *d*, H-18') and three tertiary methyl groups. In addition, a methine signal (δ 5.25, *t*, H-7) indicated a trisubstituted double bond. The foregoing evidence suggested that 1 possessed a pimaradiene-type structure and the other carbon atoms (C_3) in the molecule must be part of the acyl moiety. That the ester side chain of 1 is a malonate residue was deduced by a singlet at δ 3.43 accounting for two hydrogen atoms in its 1H NMR spectrum [1, 5]. The ^{13}C NMR spectrum of 1 (Table 1) confirmed the presence

of this residue by signals at δ 166.8 (*s*, C-1'), 40.7 (*t*, C-2') and 171.4 (*s*, C-3') [1, 6]. The remaining ^{13}C NMR absorptions were consistent with the structural features of an 7,15-pimaradiene-type skeleton with the primary ester group equatorially orientated (δ 74.7, *t*, C-18) at C-4 [7]. However, since a methine carbon (APT) occurs at an unusually upfield position (δ 38.1 *d*), a normal isopimaradiene structure for 1 was tentatively discarded [7]. Lithium aluminium hydride reduction or alkaline hydrolysis of 1 yielded the same diterpenic alcohol 2. As expected, the 1H NMR spectrum of 2 lacked resonances



- R
- 1 CH_2OCOCH_2COOH
 - 1a CH_2OCOCH_2COOMe
 - 2 CH_2OH
 - 4 CHO
 - 5 Me
 - 7 $COOH$



- R
- 3 CH_2OH
 - 6 Me

* Part 2 in the series 'Diterpenoids from *Calceolaria* species'.
 For part 1 see ref [1].

Table 1 ^{13}C NMR spectral data of compounds **1**, **1a**, **2**, **3**, **5–7** (CDCl_3 , TMS)

Carbon	1 *	1a †	2	3	5	6	7
1	36.4	36.5	36.5	36.5	36.9	36.9	35.8
2	18.0	18.3	18.2	18.2	18.9	18.9	18.0
3	37.0	37.1	36.6	36.6	43.1	43.1	37.9
4	36.5	36.5	37.5	37.5	33.1	32.9	46.2
5	38.1	38.2	37.3	37.3	43.6	43.7	38.7
6	23.8	24.0	23.7	23.7	23.8	23.8	25.2
7	118.9	119.1	119.2	118.5	119.6	119.0	118.8
8	137.0	137.0	137.1	137.7	136.9	137.4	137.0
9	53.2	53.2	53.3	53.5	53.3	53.5	53.5
10	35.0	35.1	35.0	34.9	35.2	35.3	34.7
11	25.4	25.5	25.5	25.2	25.4	25.1	25.3
12	39.4	39.5	39.6	38.2	39.7	38.2	39.6
13	39.0	39.1	39.0	35.4	38.9	35.4	38.8
14	49.6	49.7	49.7	48.8	49.7	48.8	49.7
15	146.0	145.7	145.9	28.0	145.9	28.0	145.6
16	111.4	111.4	111.3	7.8	111.1	7.7	111.3
17	30.0	30.0	29.8	28.1	29.7	27.9	29.6
18	74.7	74.5	72.6	72.5	33.5	33.5	185.4
19	18.2	18.2	18.4	18.5	22.2	22.2	17.5
20	22.6	22.7	22.8	22.8	22.7	22.7	22.5

* Malonate carbons at 166.8 (s), C-1', 40.7 (t), C-2', 171.4 ppm (s), C-3'

† Malonate carbons at 166.4 (s), C-1', 41.4 (t), C-2', 166.7 ppm (s), C-3' Methyl ester carbon at 52.4 ppm (q)

for the malonate protons and the H-18 and H-18' signals shifted upfield from δ 3.95 and 3.78 to 3.34 and 3.13, respectively.

The ^{13}C NMR data of **2** (Table 1) were very similar to the skeletal carbon absorptions of **1** with the exception of the signals near C-18. The spectrum showed, apart from other signals, a doublet (SFORD and APT) at δ 37.3 ppm which was displaced, on going from **1** to **2**, upfield by 0.8 ppm, whilst the other doublet (δ 53.3 ppm) in the spectrum appeared to be unaffected ($\Delta\delta = 0.1$ ppm). These differences could be rationalized by considering the neighbourhood effects of the acyl group on the carbon absorption at δ 38.1, which was, therefore, tentatively assigned to C-5. However, apart from the known endocyclic homoallylic effect of the Δ^7 bond [8] and the γ -gauche effect due to the C-18 oxy-methylene group, another interaction must operate in order to justify this unusually upfield absorption of C-5.

Comparison of the carbon signals tentatively assigned to C-1, C-20 and C-11 of **2** with those of isopimarol [7], shows C-1 to be shielded ($\Delta\delta = -3.1$ ppm), C-20 to be deshielded ($\Delta\delta = 6.9$ ppm) and C-11 to be deshielded ($\Delta\delta = 5.0$ ppm). These effects, which are also observed in dihydromomilactone **A** and other related compounds [9], can be explained by the interaction imposed by an uncommon orientation of C-11, which also produces an additional γ -effect on C-5, thus indicating the increased shielding of this carbon atom (compared with Dreiding molecular models). The assignments of the remaining signals in the ^{13}C NMR spectrum of **2** (Table 1), especially those of the A and C rings, were made on the basis of the observed multiplicities, empirical shift rules (α , β and γ effects) [10] and comparison with the corresponding carbon atoms of some derivatives (see Table 1). In fact, by catalytic reduction of 9-*epi-ent*-isopimarol (**2**), the vinyl group at C-13 was selectively reduced to give 15,16-

dihydro-9-*epi-ent*-isopimarol (**3**). As anticipated from a Dreiding model, the remaining double bond (Δ^7) in ring **B** was sterically hindered, resisting catalytic reduction under the usual conditions. Also as expected, the ^1H NMR spectrum of **3** lacked resonances for vinyl protons and its ^{13}C NMR spectrum clearly exhibited the characteristics of an ethyl group (28.0, *t*, C-15, 7.8, *q*, C-16) instead of a vinyl group. Treatment of 9-*epi-ent*-isopimarol (**2**) with Jones reagent [11] gave the aldehyde **4**, $\text{C}_{20}\text{H}_{34}\text{O}$ ($[\text{M}]^+$ at *m/z* 286) the spectra of which (see Experimental) are in full accordance with the assigned structure.

Huang-Minlon reduction [12] of **4** gave a mixture of two products, **5** and **6**, which were purified by chromatography on silica gel impregnated with silver nitrate. The ^1H NMR spectrum of **5** showed four methyl-group protons (C-13, C-18, C-19 and C-20), and an olefinic proton appearing at δ 5.33 was assigned to H-7 and an ABC system (δ 5.95, H-15, 4.98, H-16c, and 4.97, H-16t) was attributed to the vinylic protons.

Obviously, the replacement of the OH group at C-18 of **2** by one hydrogen atom caused the loss of a γ -gauche interaction producing, as appeared in the ^{13}C NMR spectrum of **5** (Table 1), an upfield shift of 6.3, 6.5 and 3.8 ppm on the C-5, C-3 and C-19 carbon atoms, respectively. Also, a shielding effect of 4.4 ppm on C-4 was observed. Therefore, compound **5** is shown to be 9 α -H-*ent*-7,15-isopimaradiene.

That **6** is the 15,16-dihydro derivative of **5** was evident from the ^1H and ^{13}C NMR data of this product. In fact, the ^1H NMR spectrum of **6** exhibited four tertiary methyl groups, a methine proton, and the signals due to the vinyl group were missing. In addition, all the ^{13}C NMR signals of **6** (Table 1) were in good agreement with the proposed structure. The formation of compound **6** on treatment of aldehyde **4** with hydrazine must be due, in part, to the

generation of a diimide under the reaction conditions. The diimide reduction of olefins is a well-known reaction [13]. Finally, treatment of aldehyde **4** with excess of Jones reagent gave acid **7**, the spectra of which are in full accordance with the assigned structure.

In order to remove any ambiguity concerning the structure and stereochemistry of 9 α -H-ent-isopimarol, the compound was subjected to X-ray analysis which confirmed the structure shown in **2** (Fig 1). Details of the X-ray analysis are given in the Experimental.

9-Epimeric diterpenoids constitute a relatively rare group of secondary metabolites with twelve examples known prior to the present work. These areannonalide [14], momilactones A, B, C, [8, 15], icacinol [16], icacinone [17], humirianthenolides A, B, C, D, E and F [18]. From a biogenetic point of view, the formation of these compounds might involve a chair-boat rather than a chair-chair cyclization of geranylgeranyl pyrophosphate to form a bicyclic intermediate possessing a 10 α -methyl group and a 9 α -hydrogen atom or its enantiomer, i.e. with H-9 and C-20 in a *cis* relationship. In particular, subsequent formation of ring C, elimination of a 7-hydrogen and oxidation of C-18 could afford the alcohol **2** [19]. On the other hand, the accumulation of malonyl-esters in members of the *Calceolaria* genus may be of chemotaxonomic interest.

EXPERIMENTAL

Mps uncorr, ^1H NMR 60, 100 and 400 MHz in CDCl_3 with TMS as int. standard, ^{13}C NMR 100 MHz, CDCl_3 with TMS

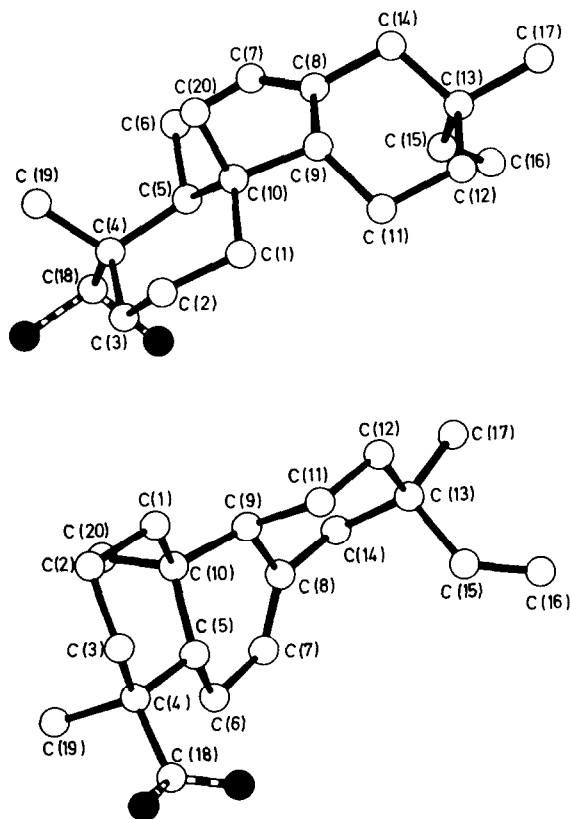


Fig 1 X-Ray crystal structure of 9-*epi-ent*-isopimarol (**2**)

as int. std. Assignments of ^{13}C NMR chemical shifts were made with the aid of APT and SFORD IR film on NaCl or KBr pellets, MS direct inlet 70 eV.

Calceolaria glandulosa Poepp. ex Benth., collected in Cuesta Zapata, V-Región, Chile, in November 1985, was identified at the Universidad Federico Santa María, where a voucher specimen is deposited.

The aerial parts of *C. glandulosa* (500 g) were extracted at room temp with CHCl_3 for 12 hr, affording 12.3 g of a syrup. This crude material was chromatographed on a silica gel column (400 g) and eluted with mixts of petrol and EtOAc of increasing polarity. Fractions of 100 ml were taken and combined based upon TLC monitoring. Fractions 5–9 gave a mixture of two unknown substances. Fractions 12–14 provided a mixt which contained a third unknown compound **1**. Fractions 16–28, contained a single substance (gummy, 6.8 g) **1**.

18-Malonyloxy-9-*epi-ent*-isopimarol (1) Viscous colourless oil, $[\alpha]_D^{25} -105.1$ (CHCl_3 , *c* 1.0). IR $\nu_{\text{max}}^{\text{film}}$ cm^{-1} 3400–3200, 3060, 2960–2840, 1740, 1715, 1450, 1375, 1310, 1150, 1000, 910, 845, 820. ^1H NMR (400 MHz) δ 5.84 (1H, *dd*, *J* = 11.2, 17.6 Hz, H-15), 5.25 (1H, *t*, *J* = 4.0 Hz, H-7), 4.96 (1H, *dd*, *J* = 11.2, 1.5 Hz, H-16c), 4.94 (1H, *dd*, *J* = 17.6, 1.5 Hz, H-16t), 3.95 (1H, *d*, *J* = 10.8 Hz, H-18), 3.78 (1H, *d*, *J* = 10.8 Hz, H-18'), 3.43 (2H, *s*, H_2 -malonyl), 1.00 (3H, *s*, Me-17), 0.97 (3H, *s*, Me-20), 0.94 (3H, *s*, Me-19). ^{13}C NMR see Table 1, MS *m/z* (rel. int.) 374 [$\text{C}_{23}\text{H}_{34}\text{O}_4$, *M*] $^+$ (2), 359 [*M* – Me] $^+$ (3), 330 [*M* – CO_2] $^+$ (1), 288 [*M* – $\text{C}_3\text{H}_5\text{O}_3$] $^+$ (7), 270 [*M* – $\text{HOOCCH}_2\text{COOH}$] $^+$ (45), 255 [270 – Me] $^+$ (76), 187 (32), 148 (50), 133 (49), 119 (70), 109 (100), 105 (73), 81 (74), 54 (77).

Methyl 18-malonyloxy-9-*epi-ent*-isopimarol (1a) After addition of $\text{CH}_2\text{N}_2\text{Et}_2\text{O}$, **1** (300 mg) was transformed to **1a**. Gummy, $[\alpha]_D^{25} -115.3$ (CHCl_3 , *c* 1.1). IR $\nu_{\text{max}}^{\text{film}}$ cm^{-1} 3080, 2960–2840, 1740, 1720, 1460, 1430, 1375, 1280, 1150, 1020, 910. ^1H NMR (100 MHz) δ 5.86 (1H, *dd*, *J* = 11.0, 17.0 Hz, H-15), 5.26 (1H, *t*, *J* = 4.0 Hz, H-7), 4.95 (1H, *dd*, *J* = 11.0, 1.5 Hz, H-16c), 5.93 (1H, *dd*, *J* = 17.0, 1.5 Hz, H-16t), 3.93 (1H, *d*, *J* = 11.0 Hz, H-18), 3.74 (1H, *d*, *J* = 11.0 Hz, H-18'), 3.72 (3H, *s*, COOMe), 3.38 (2H, *s*, H_2 -malonyl), 1.02 (3H, *s*, Me-17), 0.98 (3H, *s*, Me-20), 0.96 (3H, *s*, Me-19). ^{13}C NMR see Table 1, MS *m/z* (rel. int.) 388 [$\text{C}_{24}\text{H}_{36}\text{O}_4$, *M*] $^+$ (4), 370 (5), 270 [*M* – $\text{HO}_2\text{CCH}_2\text{CO}_2\text{Me}$] $^+$ (57), 255 [270 – Me] $^+$ (100), 251 (25), 187 (38), 148 (39), 133 (49), 119 (74), 109 (88), 105 (70), 81 (67), 55 (70).

9-*epi-ent*-isopimarol (2) **1** (1 g) was treated with LiAlH_4 in dry Et_2O . After usual work-up and crystallization from MeOH– H_2O , **2** was obtained as white crystals (700 mg). On the other hand, **1** (1 g) was also treated with K_2CO_3 in MeOH at room temp under N_2 . After 4 hr the mixt was filtered, concd and crystallized from MeOH– H_2O to yield the same compound **2** (530 mg). Mp 113–114 $^\circ$, $[\alpha]_D^{25} -168.0$ (CHCl_3 , *c* 1.0). IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} 3350, 3080, 2960–2840, 1630, 1460, 1445, 1380, 1150, 1040, 1010, 920, 845, 820. ^1H NMR (400 MHz) δ 5.89 (1H, *dd*, *J* = 10.6, 18.0 Hz, H-15), 5.26 (1H, *t*, *J* = 3.0 Hz, H-7), 4.95 (1H, *dd*, *J* = 10.6, 1.5 Hz, H-16c), 4.93 (1H, *dd*, *J* = 18.0, 1.5 Hz, H-16t), 3.34 (1H, *d*, *J* = 10.9 Hz, H-18), 3.13 (1H, *d*, *J* = 10.9 Hz, H-18'), 1.00 (3H, *s*, Me-17), 0.97 (3H, *s*, Me-20), 0.87 (3H, *s*, Me-19). ^{13}C NMR see Table 1, MS *m/z* (rel. int.) 288 [$\text{C}_{20}\text{H}_{32}\text{O}$, *M*] $^+$ (64), 273 [*M* – Me] $^+$ (58), 270 [*M* – H_2O] $^+$ (67), 257 (92), 255 (67), 241 (59), 147 (77), 134 (60), 121 (75), 109 (100), 91 (78), 55 (83), 41 (78).

15,16-Dihydro-9-*epi-ent*-isopimarol (3) **1** (200 mg) was dissolved in 100 ml of MeOH. PtO_2 (50 mg) was added and the mixt hydrogenated in a Parr apparatus at an initial press of 50 psi. After 4 hr, the mixt was filtered, concd and chromatographed on a silica gel column (10 g) and eluted with petrol–EtOAc (9/1) yielding pure **3** (160 mg). Mp 115–116 $^\circ$ (MeOH– H_2O) $[\alpha]_D^{25} -154.8$ (CHCl_3 , *c* 1.00). IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} 3320, 2960–2840,

1450, 1380, 1060, 1030, 1000, 845, 820 ^1H NMR (400 MHz) δ 5.25 (1H, *t*, $J = 4.0$ Hz, H-7), 3.36 (1H, *d*, $J = 11.0$ Hz, H-18), 3.18 (1H, *d*, $J = 11.0$ Hz, H-18'), 1.00 (3H, *s*, Me-17), 0.93 (3H, *s*, Me-20), 0.87 (3H, *s*, Me-19), 0.77 (3H, *t*, $J = 7.0$ Hz, Me-16) ^{13}C NMR see Table 1, MS m/z (rel int.) 290 [$\text{C}_{20}\text{H}_{34}\text{O}$, M] $^+$ (100), 275 [M - Me] $^+$ (86), 272 [M - H₂O] $^+$ (58), 260 (92), 259 (78), 257 (75), 243 (57), 190 (71), 187 (83), 151 (73), 147 (74), 135 (76), 133 (79), 95 (89), 93 (78), 81 (86), 55 (82), 41 (84)

9-*epi-ent-Isopimaral* (4) **1** (300 mg) was dissolved in 100 ml of Me₂CO and treated with Jones reagent for 20 min at 0°. Excess oxidant was reduced with MeOH and the products isolated with CHCl₃. This crude material was evapd and chromatographed on a silica gel column (20 g) and eluted with petrol-EtOAc (19:1) yielding pure **4** (210 mg) Mp 79–80 (MeOH-H₂O) $[\alpha]_D^{25} - 112.8$ (CHCl₃, *c* 1.00) IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹ 3080, 2960, 2840, 2680, 1720, 1630, 1450, 1380, 980, 930, 850, 820 ^1H NMR (60 MHz) δ 9.27 (1H, *s*, H-18), 5.95 (1H, *dd*, $J = 10.0, 18.0$ Hz, H-15), 5.25 (1H, *t*, $J = 4.0$ Hz, H-7), 4.96 (1H, *dd*, $J = 10.0, 1.5$ Hz, H-16 *c*), 4.93 (1H, *dd*, $J = 18.0, 1.5$ Hz, H-16 *t*), 1.10 (3H, *s*, Me-19), 0.97 (6H, *s*, Me-17, Me-20) MS m/z (rel int.) 286 [$\text{C}_{20}\text{H}_{30}\text{O}$, M] $^+$ (100), 271 [M - Me] $^+$ (22), 257 [M - CHO] $^+$ (35), 243 (30), 187 (33), 145 (22), 133 (20), 131 (33), 119 (30), 105 (39), 91 (30), 81 (34), 55 (25), 43 (18), 41 (26)

9-*epi-ent-7,15-Isopimaradiene* (5) and 9-*epi-ent-7-isopimaraene* (6) **4** (1.3 g) was treated with 2.5 g of KOH, 5 ml of 95% hydrazine hydrate and 25 ml of diethylene glycol. The mixt was refluxed for 4 hr and the condenser changed to a dist column while slow boiling was continued to removed H₂O. A white solid deposited in the column. When the pot temp had reached 240°, 2 ml of hydrazine was added and refluxing was continued for an additional 2 hr. The mixt was then cooled, acidified with concd HCl and extracted with Et₂O. The Et₂O soln and the white solid were mixed and subjected to silica gel CC (80 g, HF₂₅₄ for TLC) impregnated with AgNO₃ (10%), using *n*-pentane (1 l) and *n*-pentane-EtOAc (49:1), (2 l). Fractions of 50 ml were collected and combined based upon TLC monitoring. Fractions 12–14, containing a single compound, were mixed and afforded **6** (83 mg). Fractions 20–29, containing another pure compound, were mixed affording **5** (350 mg). **5**, white crystals, mp 73–74 $[\alpha]_D^{25} - 170.2$ (CHCl₃, *c* 1.00) IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹ 2960–2840, 1630, 1450, 1390, 1370, 1000, 920, 850, 820 ^1H NMR (400 MHz) δ 5.95 (1H, *dd*, $J = 18.0, 10.5$ Hz, H-15), 5.33 (1H, *br d*, $J = 5, 5$ Hz, H-7), 4.98 (1H, *dd*, $J = 10, 5, 1, 5$ Hz, H-16 *c*) 4.97 (1H, *dd*, $J = 18.0, 1.5$ Hz, H-16 *t*), 1.00 (3H, *s*, Me-17), 0.95 (3H, *s*, Me-18), 0.92 (3H, *s*, Me-19), 0.90 (3H, *s*, Me-20) ^{13}C NMR see Table 1, MS m/z (rel int.) 272 [$\text{C}_{20}\text{H}_{32}$, M] $^+$ (85), 257 [M - Me] $^+$ (85), 229 (34), 148 (80), 133 (77), 119 (78), 105 (100), 93 (99), 81 (91), 79 (86), 55 (92), 41 (58). Compound **6**, amorphous powder IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹ 2960–2840, 1450, 1390, 1380, 1000, 845, 820 ^1H NMR (60 MHz) δ 5.30 (1H, *t*, $J = 4.0$ Hz, H-7), 0.95 (3H, *s*, Me-18), 0.90 (9H, *br s*, Me-17, Me-19, Me-20), 0.82 (3H, *t*, $J = 7.0$ Hz, Me-16) ^{13}C NMR see Table 1, MS m/z (rel int.) 274 [$\text{C}_{20}\text{H}_{34}$, M] $^+$ (96), 259 [M - Me] $^+$ (99), 245 (84), 189 (85), 177 (64), 163 (94), 135 (94), 121 (93), 105 (100), 95 (91), 93 (94), 83 (86), 81 (92), 69 (95), 67 (90), 43 (87), 41 (86)

9-*epi-ent-7,15-Isopimaradiene-18-*oic* acid* (7) **4** (200 mg) was dissolved in 50 ml of Me₂CO and treated with Jones reagent for 1 hr at room temp. The mixt was dissolved in CHCl₃, evapd and chromatographed by silica gel CC (20 g) and eluted with petrol-EtOAc (9:1) yielding pure **7** (110 mg) Mp 170–173 (petrol-EtOAc) $[\alpha]_D^{25} - 124.5$ (CHCl₃, *c* 0.50) IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹ 3200, 3080, 2960, 2840, 1700, 1630, 1450, 1380, 1280, 1010, 915, 850, 825 ^1H NMR (400 MHz) δ 5.92 (1H, *dd*, $J = 11.0, 18.0$ Hz, H-15), 5.25 (1H, *br d*, $J = 5.5$ Hz, H-7), 4.97 (1H, *dd*, $J = 11.0, 1.5$ Hz, H-16 *c*), 4.94 (1H, *dd*, $J = 11.0, 1.5$ Hz, H-16 *t*), 4.94 (1H,

dd, $J = 18.0, 1.5$ Hz, H-16 *t*), 1.26 (3H, *s*, Me-19), 1.01 (3H, *s*, Me-17), 0.98 (3H, *s*, Me-20) ^{13}C NMR see Table 1 MS m/z (rel int.) 302 [$\text{C}_{20}\text{H}_{30}\text{O}_2$, M] $^+$ (66), 287 [M - Me] $^+$ (78), 260 (74), 259 (74), 257 (60), 241 (63), 221 (81), 187 (71), 175 (64), 133 (86), 121 (82), 119 (100), 107 (79), 105 (71), 95 (78), 81 (84), 67 (87), 55 (74), 43 (73), 41 (77)

*Crystallographic data for 9-*epi-ent-isopimarol* (2)* A prismatic crystal (0.6 × 0.4 × 0.2 mm) was mounted on an automatic four-circle diffractometer (Philips PW 1100), using CuK α radiation. The system was monoclinic, and the data collected assuming a C2 space group $a = 28.812$ (12), $b = 7.351$ (4), $c = 17.882$ (8) Å and $\beta = 112.08$ (8). The unit cell contains eight molecules. Out of 3420 measured reflections, only 2831 were considered as observed [$I > 3\sigma(I)$]. The structure was solved by direct methods [20] and the parameters refined to an R value of 5.8% [21]. The two independent molecules of the asymmetric unit appear on Fig. 1. In both, the hydroxy group takes two positions, occupied equally, corresponding to two hydrogen bonding schemes. The relative configuration is obviously the same and is represented by the formula **2**.

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